Synthesis of Water-soluble Sugar Derivatives of Combretastatin A-4

Richard T. Brown,^{*,a} Brian W. Fox,^b John A. Hadfield,^b Alan T. McGown,^b Stephen P. Mayalarp,^{*,a,b} George R. Pettit^c and Julie A. Woods^b ^a Department of Chemistry, University of Manchester, Manchester M13 9PL, UK ^b CRC Department of Experimental Chemotherapy, Paterson Institute for Cancer Research, Christie Hospital NHS Trust, Wilmslow Road, Manchester M20 9BX, UK

° Cancer Research Institute and Department of Chemistry, Arizona State University, Tempe, Arizona 85287-1604, USA

Three sugar derivatives (6, 7 and 13) of the antimitotic agent combretastatin A-4 1 have been synthesized. Two of these water-soluble derivatives (6, 13) showed strong cytotoxicity to the murine P388 cell line.

Combretastatin A-4 1, a natural product isolated from *Combretum caffrum*, is currently being investigated under the sponsorship of the Cancer Research Campaign Clinical Trials scheme. This stilbene 1 has been shown to be one of the most potent inhibitors of tubulin assembly yet discovered. A close structural analogue, Combretastatin A-1 2, has also been shown to alter tubulin dynamics, a mechanism of action common to the clinically useful *Vinca* alkaloid class of anti-cancer agents. These agents are known to belong to the group of compounds that show extensive cross-resistance in multidrug-resistance (MDR) cell lines.

A major problem encountered with combretastatin A-41 has been its insolubility. So far no suitable formulation has been developed so as to allow the commencement of clinical trials. Herein we describe the synthesis of three water-soluble derivatives of combretastatin A-41.



Structures of compounds 1-3

Drugs such as morphine and paracetamol conjugate to glucuronic acids *in vivo* to afford paracetamol β -D-glucuronide $3^{1,2}$ and morphine 3- and 6- β -D-glucuronide. 3^{-6} Morphine 6-glucuronide is now thought to be the active analgesic.⁷ The methodology developed^{8,9} to synthesize the conjugate glycoside 3 via C-1 imidate 4 was used to prepare β -D-melibioside and β -D-glucuronide derivatives of combretastatin A-4 1 whilst a protected bromoglucose was used to synthesize a combretastatin β -D-glucoside.

Results and Discussion

Methyl (trichloroacetimidoyl 2,3,4-tri-O-isobutyryl- α -D-glucopyranosid)uronate **4** was allowed to react with the phenol **1** in the presence of boron trifluoride-diethyl ether to afford the protected combretastatin glucuronide **5** in 68% yield. Deprotection of compound **5** using aq. sodium hydroxide, followed by chromatography on Sephadex G15, gave the required combretastatin β -D-glucuronide **6** as a chromatographically pure gum in 77% yield (Scheme 1).

Combretastatin melibioside 7 was prepared in a similar fashion from a melibiose C-1 imidate 8. Briefly, melibiose 9 was treated with isobutyryl chloride in the presence of pyridine to



Scheme 1 Synthesis of combretastatin glucuronide 6. DCM = dichloromethane. *Reagents:* i, 1 + BF₃·Et₂O, DCM; ii, NaOH, MeOH.

give the octaisobutyryl ester of melibiose, compound 10, as an α/β mixture (6:1 from NMR analysis) in excellent yield. Selective removal of the ester from the anomeric hydroxy group was achieved by treatment of the octaisobutyryl ester 10 with gaseous ammonia, and afforded the heptaisobutyrate of the hemiacetal, compound 11, in 64% yield. Treatment of the hemiacetal 11 with trichloroacetonitrile followed by filtration through silica gel gave the ester-protected melibiose C-1 imidate **8** as a mixture of α/β anomers in excellent yield. On account of its complexity the NMR spectrum of compound 8 could not be fully characterised but showed the expected shift in the position of the C-1 anomeric proton. Reaction of combretastatin A-41 with imidate 8 in the presence of boron trifluoride-diethyl ether yielded the desired combretastatin melibioside 12 as indicated by fast-atom bombardment (FAB) mass spectrometry and ¹H NMR spectroscopy. Saponification of heptaester 12 with aq. sodium hydroxide, followed by chromatography on Sephadex G15, afforded combretastatin melibioside 7 as a water-soluble gum (Scheme 2)

Combretastatin glucoside 13 was prepared from glucose in



Scheme 2 Synthesis of combretastatin melibioside 7. DCM = dichloromethane, Py = pyridine. Reagents: i, $Pr^{i}COCl$, Py, DCM; ii, NH_{3} , DCM, MeOH; iii, $CCl_{3}CN$, $K_{2}CO_{3}$, DCM; iv, $1 + BF_{3}\cdot Et_{2}O$; v, NaOH, MeOH.



Schemes 3 and 4 Synthesis of combretastatin glucoside 13. DCM = dichloromethane, AcOH = acetic acid. *Reagents:* i, 30% HBr-AcOH, DCM; ii, 1 mol equiv. of 5% LiOH, MeOH; iii, 5 mol equiv. of 5% aq. LiOH, MeOH; iv, CCl₃CN, K₂CO₃, DCM; v, BF₃•Et₂O; vi, LiOH

Table 1 Effect of compounds **1**, **6**, **7** and **13** on the growth of P388 mouse leukaemia cells, the multidrug resistant subline P388PR8/22, A2780 human ovarian tumour cells and the multidrug resistant subline A2780/ADR. Values shown are the concentrations (μ mol dm⁻³) required to cause a 50% decrease in cell growth.

	Cell lines			
	P388	P388PR/22	A2780	A2780/ADR
1	0.002 6	0.001 2	0.000 72	0.000 84
6	0.28	nd	nd	nd
7	> 50	> 50	> 50	> 50
13	3.8	8.3	12.8	> 50

nd = not determined.

four steps as follows. Pentaisobutyrylglucose 14, prepared in high yield from D-glucose and isobutyryl chloride, was treated with 30% hydrogen bromide in acetic acid to give the α -bromo sugar 15. Coupling of combretastatin A-4 1 with the bromoglucose 15 with 1 mol equiv. of lithium hydroxide in methanol followed by hydrolysis with 5 mol equiv. of lithium hydroxide produced the desired combretastatin β -D-glucoside 13 in 80% yield (Scheme 3). This glucoside 13 was also prepared using the imidate chemistry as described above (Scheme 4). Briefly, ammonolysis of pentaester 14 yielded an α/β anomer mixture of the tetraester hemiacetal 16 which was converted into the imidate 17 in high yield. Acid-catalysed coupling of imidate 17 with the phenol 1 gave the ether 18, which afforded combretastatin β -D-glucoside 13 on hydrolysis.

All three combretastatin sugar derivatives (6, 7, 13) were tested for *in vitro* cytotoxicity in a P388 murine leukaemia cell line. The melibioside 7 and the glucuronide 6 were also tested in the P388PR8/22 MDR and in the A2780 human ovarian cell lines. Two of the sugar derivatives (6, 13) showed cytotoxic activity (Table 1) but were not as active as the parent phenol 1.

Experimental

All solvent extracts of aqueous solutions were dried over anhydrous sodium sulfate. Hexanes refer to a light petroleum fraction boiling at 40–60 °C. NMR spectra were recorded on Bruker AC300 or Varian Unity 500 spectrometers and are referenced to tetramethylsilane. J Values are given in Hz. Optical rotations were measured on a Optical Activity AA-100 polarimeter; $[\alpha]_D$ -values are given in units of 10^{-1} deg cm² g⁻¹. Mass spectra were recorded on a Kratos Concept mass spectrometer at an ionising energy of 70 eV; for CI spectra ammonia was the reagent gas, and for FAB spectra the matrix used was 3-nitrobenzyl alcohol. IR spectra were determined on a Perkin-Elmer 1710 FT spectrometer. UV spectra were determined on a Shimadzu UV-260 spectrophotometer. Mps were measured on a Kofler block and are uncorrected. TLC was carried out on plates coated with silica $60F_{254}$.

Methyl (Combretastatinyl 2,3,4-Tri-O-isobutyryl-B-D-glucopyranosid)uronate 5.---To a solution of combretastatin A-4 1 (500 mg, 1.58 mmol) and the imidate 4 (178 mg, 3.17 mmol) in anhydrous dichloromethane (15 cm³) at <0 °C (ice–NaCl) under N_2 was added boron trifluoride-diethyl ether (400 mm³, 3.17 mmol) and the mixture allowed to warm to room temperature and stirred for 3 days. The resulting red solution was diluted with dichloromethane, washed successively with saturated aq. sodium hydrogen carbonate and brine, and dried, and the solvent was removed under reduced pressure. The glucuronate 5 was isolated as a solid (770 mg, 68%) after chromatography on silica gel with dichloromethane-ethyl acetate (9:1) as eluent. Trituration with dichloromethane afforded the *title ester* 5 as needles, mp 127–131 °C; R_f 0.55 [dichloromethane-ethyl acetate (9:1)]; $v_{max}(film)/cm^{-1}$ 3372, 3320, 3246, 1694 and 1609; λ_{max} (EtOH)/nm 230, 244, 288 and 314 (no base shift on addition of NaOH); $\delta_{\rm H}$ (CDCl₃-D₂O) 7.04 (1 H, d, J 2, 2'-H), 7.00 (1 H, dd, J 8 and 2, 6'-H), 6.76 (1 H, d, J 8, 5'-H), 6.49 (2 H, s, 2- and 6-H), 6.48 (1 H, d, J 12, 1a-H), 6.46 (1 H, d, J 12, 1a-H), 5.39-5.25 (3 H, m, 2"-, 3"- and 4"-H), 4.94 (1 H, d, J 7, 1"-H), 3.97 (1 H, d, J 8, 5"-H), 3.97, 3.78, 3.71 and 3.70 (15 H, 4 \times s, 4 \times OMe, 1 \times CO₂Me), 2.58–2.42 (3 H, m, CHMe₂) and 1.37-0.85 (18 H, m, CHMe₂) [(FAB) M⁺, 716.3044. C₃₇H₄₈O₁₄ requires M, 716.3044].

Combretastatinyl B-D-Glucopyranosiduronic Acid Sodium Salt 6.--To a solution of combretastatin glucuronate 5 (0.73 g, 1.02 mmol) in methanol (18 cm³) was added 5% aq. sodium hydroxide (3.26 cm^3) and the mixture was stirred overnight. Removal of the solvent under reduced pressure, followed by chromatography on Sephadex G15 with water as eluent, afforded the sodium salt 6 (424 mg) as a yellow solid containing a trace amount of sodium isobutyrate (NMR spectroscopy); $R_{\rm f}$ 0.59 [ethyl acetate-methanol-water-acetic acid (60:30:9:1)]; λ_{max} (water)/nm 240 and 288 (no base shift with NaOH); δ_H(D₂O-[²H₆]acetone) 7.17 (1 H, d, J 2, 2'-H), 7.00 (1 H, dd, J 8 and 2, 6'-H), 6.96 (1 H, d, J 8, 5'-H), 6.62 (2 H, s, 2- and 6-H), 6.59 (1 H, d, J 12, 1a-H), 6.52 (1 H, d, J 12, 1'a-H), 4.76 (1 H, d, J 7.5, 1"-H), 3.88, 3.79 and 3.71 (12 H, $3 \times s$, $4 \times OMe$) [(FAB) 537.1341 (M⁺ + Na). $C_{24}H_{27}Na_2O_{11}$ requires m/z537.1349].

Melibiose Octaisobutyrate 10.—To a vigorously stirred suspension of α -melibiose hydrate 9 (5.0 g, 13.9 mmol) in a mixture of pyridine (12 cm³) and dichloromethane (30 cm³) at -8 °C (ice–NaCl) was added isobutyryl chloride (17.3 cm³, 0.167 mol) over a period of 35 min whilst the temperature of the mixture during this very exothermic reaction was kept below 15 °C. The mixture was then allowed to warm to room temperature and was stirred overnight. The resulting suspension was diluted with dichloromethane, washed successively with 1 mol dm⁻³ HCl, saturated aq. sodium hydrogen carbonate and brine, and dried and the solvent was removed under reduced pressure to leave a pale yellow oil. Trituration with hexanes afforded the crude product **10** as a solid (15.4 g) as an α/β anomeric mixture (6:1, by NMR spectroscopy); R_f 0.60 [hexane-ethyl acetate (3:1)]; $\delta_{\rm H}({\rm CDCl}_3)$ 6.28 (1 H, d, J 3.5, 1-H^{lpha}), 5.69 (1 H, d, J 8, 1-H^{eta}), 5.54 (1 H, t, J 10, 3-H), 5.50 (1 H, d, J 3, 1'-H), 5.41 (1 H, dd, J 10 and 3, 2'-H), 5.23 (1 H, t, J 10, 4-H), 5.13-5.07 (1 H, m, 3'-H), 5.04 (1 H, dd, J 10 and 3.5, 2-H), 4.25-4.15 (2 H, m, 6-H₂), 4.07-3.95 (2 H, m, 5- and 5'-H), 3.69 (1 H, dd, J 12 and 4.5, 6'-H), 3.48 (1 H, dd, J 12 and 1.5, 6'-H), 2.72-2.35 (8 H, m, CHMe₂) and 1.25-1.06 (48 H, m, CHMe₂); m/z (FAB) 901 (M – H).

Heptaisobutyrylmelibiose Imidate 8.—Into a solution of the crude melibiose octaisobutyrate 10 (12.0 g) in a mixture of dichloromethane (20 cm³) and methanol (20 cm³) at -8 °C (ice–NaCl) was bubbled gaseous ammonia during 20 min. The mixture was stirred at room temperature for 18 h, more ammonia was passed through for 30 min, and the mixture was stirred for a further 30 min. Evaporation off of the solvent and chromatography of the residue on silica gel with hexanes-ethyl acetate (3:1) as eluent afforded the heptaisobutyrate 11 as an oil (7.4 g, 64%), R_f 0.38 [hexanes-ethyl acetate (3:1)]; $v_{max}(film)/cm^{-1}$ 3474 (OH).

To a solution of the hemiacetal **11** (7.4 g, 8.9 mmol) in anhydrous dichloromethane (85 cm³) were added trichloroacetonitrile (2.23 cm³, 22.3 mmol) and anhydrous potassium carbonate (3.7 g, 26.7 mmol) and the mixture was stirred overnight. The solution was filtered through a pad of silica gel and the product was isolated by elution with diethyl ether (250 cm³). Evaporation off of the solvent afforded the gummy imidate **8** as an α/β (3:2) mixture (7.8 g, 89%), $R_{\rm f}$ 0.46 and 0.59 [hexanes-ethyl acetate (3:1)]; $\delta_{\rm H}$ (CDCl₃) 9.22 [1 H, s, NH (β)], 8.88 [1 H, s, NH (α)], 6.47 (1 H, d, J 3.5, 1-H^{α}) and 5.84 (1 H, d, J 8, 1-H^{β}).

Heptaisobutyryl Ester of Combretastatin Melibioside 12.— To a solution of combretastatin A-41 (100 mg, 0.316 mmol) and melibiose imidate 8 (1.24 g, 1.266 mmol) in dichloromethane (5 cm³) in the dark at <0 °C (ice–NaCl) under nitrogen was added boron trifluoride–diethyl ether (40 mm³, 0.316 mmol) and the mixture was allowed to warm to room temperature overnight. The resulting red solution was diluted with dichloromethane, washed successively with saturated aq. sodium hydrogen carbonate and brine, and dried, and the solvent was removed under reduced pressure to give compound 12 R_f 0.35 [hexanes– ethyl acetate (3:1)], δ_H 6.97 (1 H, d, J 2, 2'-H), 6.94 (1 H, dd, J9 and 2, 6'-H), 6.74 (1 H, d, J9, 5'-H), 6.51 (2 H, s, 2- and 6-H), 6.46 (1 H, d, J 12, 1a-H), 6.43 (1 H, d, J 12, 1'a-H) and 5.60 (1 H, t, J 10, 3"-H) [(FAB) M⁺, 1130.5337. C₅₈H₈₂O₂₂ requires M, 1130.5298].

Combretastatin Melibioside 7.—To a solution of the crude protected combretastatin melibioside 12 (681 mg) in methanol (17 cm³) cooled in ice-salt was added 5% aq. sodium hydroxide (3.38 cm³) and the mixture was allowed to warm to room temperature overnight. Removal of the solvent under reduced pressure, followed by chromatography on Sephadex G15 with water as eluent, afforded combretastatin melibioside 7 as a yellow gum (382 mg), λ_{max} (water)/nm 238 and 290 (no base shift on addition of NaOH); δ_{H} (D₂O) 6.98 (1 H, d, J 2, 2'-H), 6.88 (1 H, dd, J 8 and 2, 6'-H), 6.82 (1 H, d, J 8, 5'-H), 6.62 (1 H, d, J 12, 1a-H), 6.60 (2 H, s, 2- and 6-H), 6.55 (1 H, d, J 12, 1'a-H), 4.85 (1 H, d, J 3, 1'''-H), 4.40 (1 H, d, J 8, 1''-H) and 3.81, 3.74 and 3.65 (12 H, 3 × s, 4 × OMe) [(FAB) 663.2220 (M + Na⁺). C₃₀H₄₀NaO₁₅ requires *m*/*z* 663.2265].

1,2,3,4,6-*Penta*-O-*isobutyryl*-D-glucopyranose 14.—A suspension of D-glucose (48.0 g, 0.267 mol) in pyridine (200 cm³) was heated on a steam-bath for 1.5 h and then was allowed to cool to $-7 \degree$ C (ice–NaCl–MeOH). To a mechanically stirred suspen-

sion of the above mixture was added a solution of isobutyryl chloride (180 cm³) in dichloromethane (200 cm³) over a period of 4.5 h at such a rate as to keep the reaction temperature below 10 °C. After half of the isobutyryl chloride-dichloromethane solution had been added, the reaction mixture was diluted with dichloromethane (200 cm³). When the addition was complete, the mixture was diluted with dichloromethane (600 cm^3), washed successively with 1 mol dm⁻³ HCl (3×200 cm³) and saturated aq. sodium hydrogen carbonate ($3 \times 200 \text{ cm}^3$). The organic phase was separated and added to 0.880 aq. ammonia (200 cm³). The two-phase mixture was stirred for 20 min and the aqueous layer was decanted. The dichloromethane solution was washed with brine, dried, and filtered, and the solvent was removed under reduced pressure to leave a yellow gum which solidified. Recrystallisation from ethanol afforded a 1:1 α/β mixture of glucose pentaisobutyrate 14 as crystals (119 g, 84%), mp 52-54 °C (Found: C, 59.1; H, 8.0. C₂₆H₄₂O₁₁ requires C, 58.9; H, 7.90%); v_{max} (film)/cm⁻¹ 2976, 2879, 1754, 1472 and 1389; m/z (CI) 548 (M + 18). Both anomers can be separated by chromatography on silica gel with hexanes-ethyl acetate (3:1) as eluent: α -anomer, mp 76–78 °C (from EtOH); $[\alpha]_D$ + 70 (c 28 mg cm⁻³, CHCl₃); $\delta_{\rm H}$ (CDCl₃) 6.60 (1 H, d, J 4, 1-H), 5.61 (1 H, t, J 9.5, 3-H), 5.20 (1 H, t, J 9.5, 4-H), 4.83 (1 H, dd, J 9.5 and 4, 2-H), 4.31 (1 H, ddd, J 9.5, 4.8 and 2.4, 5-H); 4.26 (1 H, dd, J 11 and 4.8, 6-H), 4.14 (1 H, dd, J 11 and 2.4, 6-H), 2.65-2.41 (5 H, m, CHMe₂) and 1.20-1.05 (30 H, m, CHMe₂); β-anomer, mp 106 °C (from EtOH); $[\alpha]_{D}$ +40 (c 32 mg cm⁻³ CHCl₃); δ_H(CDCl₃) 5.73 (1 H, d, J 8, 1-H), 5.32 (1 H, t, J 9.5, 3-H), 5.18 (1 H, dd, J 9.5 and 8, 2-H), 5.17 (1 H, t, J 9.5, 4-H), 4.23 (1 H, dd, J12.5 and 4.5, 6-H), 4.11 (1 H, dd, J12.5 and 2, 6-H), 3.85 (1 H, ddd, J 9.5, 4.5 and 2, 5-H), 2.65–2.42 (5 H, m, 5 × CHMe₂) and 1.18 (30 H, m, $5 \times \text{CHMe}_2$).

2,3,4,6-Tetra-O-isobutyryl-a-D-glucopyranosyl Bromide 15.— To a solution of 1,2,3,4,6-penta-O-isobutyryl-D-glucopyranose $(1:1 \alpha/\beta \text{ mixture})$ 14 (5.0 g, 9.4 mmol) in dichloromethane (10 cm³) at < 0 °C (ice-NaCl) was added 30% hydrogen bromide in acetic acid (10 cm³). After warming to room temperature overnight, the resulting solution was evaporated to dryness under reduced pressure. The residue was redissolved in dichloromethane, washed successively with cold, saturated aq. sodium hydrogen carbonate and brine, and dried, and the solvent was removed under reduced pressure. Recrystallisation from ethanol afforded the α -bromide 15 as crystals (2.46 g, 50%), mp 67–69 °C; $R_f 0.8$ [CHCl₃–MeOH (9:1)]; $[\alpha]_D$ +189 (c 17 mg cm⁻³, CHCl₃) (Found: C, 50.5; H, 6.9; Br, 15.3. C₂₂H₃₅BrO₉ requires C, 50.6; H, 6.7; Br, 15.1%); v_{max}(film)/cm⁻¹ 2976, 1747, 1471, 1390 and 1248; $\delta_{\rm H}$ (CDCl₃) 6.61 (1 H, d, J 4, 1-H), 5.62 (1 H, t, J 10, 3-H), 5.21 (1 H, t, J 10, 4-H), 4.84 (1 H, dd, J 10 and 4, 2-H), 4.39-4.23 (2 H, m, 5- and 6-H), 4.15 (1 H, dd, J 12.5 and 1.5, 6-H), 2.68–2.42 (4 H, m, 4 × CHMe₂) and 1.21–0.97 (24 H, m, 4 × CHMe₂); m/z (FAB) 523 (M + H), 443, 373, 285, 197 and 71.

2,3,4,6-*Tetra*-O-*isobutyryl*-D-glucopyranose 16.—Into a solution of D-glucose pentaisobutyrate 14 (1:1 α/β mixture) (57.46 g, 0.108 mol) in a mixture of dichloromethane (1.2 dm³) and MeOH (800 cm³) cooled in ice-salt was passed gaseous NH₃ over a period of 4 h and the mixture was left to react overnight at ambient temperature. On each of the following days, more gaseous NH₃ was bubbled through for 2–3 h. The solvent was then removed under reduced pressure, the residue was redissolved in dichloromethane, the solution was washed successively with water and brine, and dried, and the solvent was evaporated off. The crude mixture was chromatographed on silica gel with hexanes–ethyl acetate (2:1) as eluent to give the tetra ester 16 as a gum (14.0 g, 28%) as an α/β mixture of hemiacetals. Trituration with hexanes gave the pure α -

hemiacetal, mp 85–87 °C; $[\alpha]_D$ + 38 (*c* 15 mg cm⁻³, CHCl₃); *R*_f 0.38 [hexanes–ethyl acetate (2:1)]; *v*_{max}(film)/cm⁻¹ 3469, 2976, 1749, 1472, 1390 and 1251; δ_H (CDCl₃) 5.60 (1 H, t, *J* 10, 3-H), 5.46 (1 H, d, *J* 4, 1-H^{α}), 5.31 (1 H, d, *J* 9, 1-H^{β}), 5.14 (1 H, t, *J* 10, 4-H), 4.91 (1 H, dd, *J* 10 and 4, 2-H), 4.28 (1 H, ddd, *J* 10, 4 and 2, 5-H), 4.21–4.10 (2 H, m, 6-H₂), 2.69–2.37 (4 H, m, 4 × CHMe₂) and 1.30–1.04 (24 H, m, 4 × CHMe₂); *m*/*z* (EI) 460, 443, 373, 355, 285 and 197; *m*/*z* (CI) 478 (M + 18).

2,3,4,6-Tetra-O-isobutyryl-a-D-glucopyranosyl Trichloro-

acetimidate 17.—To a solution of the α/β hemiacetal 16 (3.74 g, 8.13 mmol) in dichloromethane (45 cm^3) at room temperature was added trichloroacetonitrile (2.46 cm³, 3.52 g, 24.39 mmol) and the solution was stirred for 10 min. Anhydrous potassium carbonate (3.93 g, 28.46 mmol) was added and 20 h later the mixture was filtered through a pad of silica gel and the product was eluted with diethyl ether. The filtrate was concentrated under reduced pressure to afford a mixture of α/β anomers (1:1 by NMR spectroscopy) of the imidate 17 as a gum (4.79 g, 98%), $R_{\rm f}$ 0.71 and 0.64 [hexanes-ethyl acetate (2:1)]; $v_{\rm max}$ (film)/cm⁻¹ 3300, 2976, 1749, 1679, 1471, 1388 and 1248; $\delta_{\rm H}(\rm CDCl_3)$ 8.79 and 8.75 (2 H, 2 × s, 2 × NH), 6.27 (1 H, d, J4, 1-H), 5.32 (1 H, dd, J 10 and 9.5, 3-H), 5.17 (1 H, t, J 10, 4-H), 5.16 (1 H, dd, J 9.5 and 4, 2-H), 4.24-4.13 (2 H, m, 6-H), 4.07 (1 H, ddd, J10, 5 and 2, 5-H), 2.67–2.35 (4 H, m, $4 \times CHMe_2$) and 1.27–1.00 (24 H, m, $4 \times \text{CH}Me_2$; m/z (EI) 460, 443, 355 and 286; m/z (CI) 588, 568, 478, 443 and 355.

Combretastatinyl 2,3,4,6-Tetra-O-isobutyryl- β -D-glucopyranoside 18.-To a solution of combretastatin A-4 1 (200 mg, 0.63 mmol) and the glucose imidate 17 (2.296 g, 3.8 mmol) in dichloromethane (10 cm³), at room temperature under nitrogen, was added boron trifluoride-diethyl ether (477 mm³, 3.8 mmol) and the mixture was stirred overnight in the dark. The resulting purple solution was diluted with dichloromethane, washed successively with saturated aq. sodium hydrogen carbonate and brine, and dried, and the solvent was removed under reduced pressure. Chromatography on silica gel with dichloromethane-ethyl acetate as eluent (20:1 to 8:1) gave the protected glucoside 18 as a solid (435 mg, 91%), mp 193–198 °C; R_f 0.85 [dichloromethane-ethyl acetate (4:1)]; $v_{max}(film)/cm^{-1}$ 2975, 2939, 2877 and 1749; $\lambda_{max}(EtOH)/nm$ 324, 240 and 225 (no base shift on addition of NaOH); $\delta_{\rm H}$ 7.09 (1 H, d, J 2, 2'-H); 7.02 (1 H, dd, J 2 and 8, 6'-H), 6.79 (1 H, d, J 8, 5'-H), 6.54 (2 H, s, 2- and 6-H), 6.49 (1 H, d, J 12, 1a-H), 6.46 (1 H, d, J 12, 1'a-H), 5.30 (1 H, t, J 9.5, 3"-H), 5.16 (1 H, t, J 9.5, 4"-H), 5.00 (1 H, dd, J 9.5 and 8, 2"-H), 4.86 (1 H, d, J 8, 1"-H), 4.23 (1 H, dd, J 12 and 4.5, 6"-H), 4.16 (1 H, dd, J 12 and 2, 6'-H), 3.96, 3.91 and 3.86 (12 H, $3 \times s$, $4 \times OMe$), 3.80–3.70 (1 H, m, 5"-H), 2.68–2.42 (4 H, m, $4 \times CHMe_2$) and 1.25–1.08 (24 H, m, 4 × CHMe₂); m/z 758 (M⁺), 443 and 316.

Combretastatin Glucoside 13 .--- To a solution of combretastatin A-4 1 (205 mg, 0.65 mmol) in 2.4% methanolic lithium hydroxide (1.4 cm³) was added the bromo sugar 15 (0.339 g, 0.65 mmol). After 2.5 h more methanolic lithium hydroxide (1.4 cm³) was added and the mixture was stirred for 16 h. Another aliquot of methanolic lithium hydroxide (4.2 cm³) was added, the mixture was left for 4 h, and the solvent was removed under reduced pressure. The mixture was separated on Sephadex G-10 with water as eluent to afford combretastatin glucoside 13 as a solid (249 mg, 80%), mp 92-95 °C (Found: C, 58.1; H, 6.3. $C_{24}H_{30}O_{10}H_2O$ requires C, 58.1; H, 6.5%); R_f 0.54 [ethyl acetate-methanol-water-acetic acid (60:30:9:1)]; $\hat{\lambda}_{max}(water)/(1-2)$ nm 220, 240 and 286 (no shift on addition of NaOH); $\delta_{\rm H}({\rm D_2O})$ 6.97 (1 H, br s, 2'-H), 6.96 (1 H, dd, J 8 and 2, 6'-H), 6.91 (1 H, br s, 5'-H), 6.58 (2 H, br s, 2- and 6-H), 6.56 (1 H, d, J 12, 1a-H), 6.51 (1 H, d, J 12, 1'a-H), 4.40 (1 H, d, J 8, 1"-H) and 3.79, 3.71 and 3.63 (12 H, 3 \times s, 4 \times OMe) [(FAB) M⁺, 478.1846. $C_{24}H_{30}O_{10}$ requires M, 478.1839].

Cytotoxicity testing was performed by using an MTT assay.¹⁰

Acknowledgements

We thank the Cancer Research Campaign for supporting this work.

References

- B. B. Brodie and J. Axelrod, J. Pharmacol., 1948, 94, 29.
 B. B. Brodie and J. Axelrod, J. Pharmacol., 1949, 97, 58.
- 3 L. A. Woods, J. Pharmacol. Exp. Ther., 1954, 112, 158.
- 4 J. M. Fujimoto and E. L. Way, J. Pharmacol. Exp. Ther., 1957, 121, 340.

- 6 K. Yoshimura, K. Oguri and H. Tsukamoto, Chem. Pharm. Bull., 1968, **16**, 2114.
- 7 R. Osborne, P. Thompson, S. Joel, D. Trew, N. Patel and M. Slevin, Br. J. Clin. Pharmacol., 1992, 34, 130. 8 F. Scheinmann, K. W. Lumbard, R. T. Brown and S. P. Mayalarp,
- Int. Pat., 1993, WO 93/3051.
- 9 R. T. Brown, S. P. Mayalarp, A. T. McGown and J. A. Hadfield, J. Chem. Res. (S), 1993, 496.
- 10 J. M. Edmondson, L. S. Armstrong and A. O. Martinez, J. Tissue Culture Methods, 1988, 11, 15.

Paper 4/05733C Received 20th September 1994 Accepted 31st October 1994